

Fig. 1.—Observed second-order rate constants vs. $(1/[I])$ for the oxidation of methionine with iodine at pH 7.2. Initial $C_6H_{11}NO_2S = 6.82 \times 10^{-4} M$; initial total $I_2 = 10^{-4} M$; phosphate buffer = $0.02 M$; ionic strength = 2.05 (adjusted with KCl); temperature = 25.0° .

tration of free iodine is inversely proportional to that of iodide at higher concentrations.

In Fig. 2 the product of observed second-order rate constants obtained in the above fashion and the iodide concentration for each run have been plotted as a function of the pH of the system. Different iodide concentrations were used to adjust the observed rate to a convenient range. From the profile, it is evident that the rate is essentially inversely proportional to the hydrogen concentration in the range between pH 4 and 8. Above pH 8 the curve levels off, the observed second-order rate constant at pH 10 being approximately twice that at pH 9. Since the pK_{a2} of methionine in this system is approximately 9, the observed pH dependence strongly suggests that the free amino group is involved in this reaction.

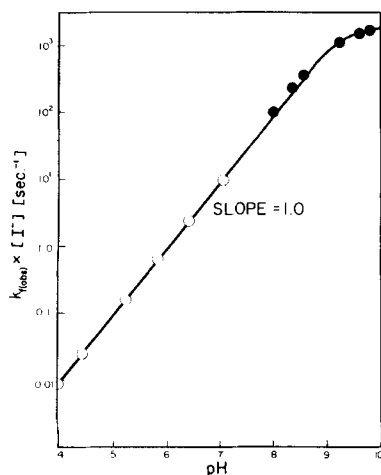


Fig. 2.—Products of observed second-order rate constant and iodide concentration vs. pH for the oxidation of methionine with iodine. Initial $C_6H_{11}NO_2S = 5 \times 10^{-4} M$; initial total $I_2 = 10^{-4} M$; phosphate or phthalate buffer = $0.02 M$; $\circ = k_{f(obs.)}$ measured in $0.1 M$ KI, $\bullet = k_{f(obs.)}$ measured in $1 M$ KI; temperature = 25.0° .

On the basis of the results presented in Figs. 1 and 2 the following rate law can be written:

$$\begin{aligned} \frac{-d[I_2]_T}{dt} &= k_{f(obs.)} [Me]_T [I_2]_T \\ &= k_f [Me^-] [I_2] \quad (\text{Eq. 4}) \end{aligned}$$

where Me^- represents the anionic methionine species and k_f is the second-order rate constant for the species Me^- and I_2 of the forward reaction. Considering Eq. 2 and the dissociation of methionine as expressed by

$$K_{a2} = \frac{[Me^-] [H^+]}{[Me]} \quad (\text{Eq. 5})$$

we can write the forward rate as

$$\frac{-d[I_2]_T}{dt} = \frac{k_f K_{a2} [Me]_T [I_2]_T}{([H^+] + K_{a2})(1 + K_1[I^-])} \quad (\text{Eq. 6})$$

and for the observed second-order rate constant as follows

$$k_{f(obs.)} = \frac{k_f K_{a2}}{([H^+] + K_{a2})(1 + K_1[I^-])} \quad (\text{Eq. 7})$$

From Eq. 7 and the data as shown in Figs. 1 and 2, a mean value for k_f at 25° was determined to be $6.5 \times 10^6 M^{-1} \text{sec}^{-1}$.

Rate of Reduction of Dehydromethionine.—

Reverse Reaction.—The reduction of the cyclic species by iodide was readily followed spectrophotometrically by the appearance of the I_3^- species. Pseudo first-order rate constants, for the reverse reaction, $k_r(obs.)$, were obtained from such data in either of two ways. For fast reaction at constant pH and excess iodide concentration, pseudo first-order rate constants were obtained by plotting $\log(A_\infty - A)$ against time. If the rate was expected to be slow, a relatively high concentration of dehydromethionine was used, and the initial rate of formation was measured. Based on previously determined values for the molar absorptivity of iodine in the particular system, the initial rate of reaction was converted to pseudo first-order rate constants by dividing through by the initial dehydromethionine concentration.

Pseudo first-order rate constants obtained in this fashion at different pH values are shown plotted versus pH in Fig. 3. From the resulting pH profile it is evident that above pH 4.5 the rate is directly proportional to the hydrogen-ion concentration, while in the more acidic region the order of the over-all reaction with respect to the hydrogen concentration is larger than 1.

The dependence of the reverse pseudo first-order rate constant on the iodide concentration at pH of 5.02 is evident from the log-log data plotted in Fig. 4. The line which closely fits the experimental points was drawn with a slope of 2. On this basis, it would appear that the kinetic dependence with respect to iodide is 2. Similar determinations at pH 2.48, however, yielded a log-log plot with a slope of 1.52 also shown in Fig. 4. From these studies it would appear that the responsible reaction pathway in the acidic region may differ at least in part from that operating at neutral pH. If we assume that the protonated dehydromethionine requires an attack by only a single iodide anion in the rate-determining step while the

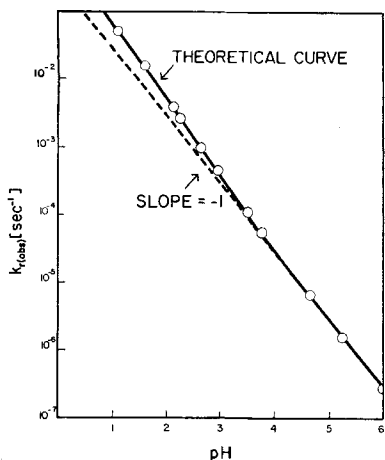


Fig. 3.—Observed first-order rate constant vs. pH for the reduction of dehydromethionine. KI = 0.1 M; C₅H₉NO₂S = 8.3 × 10⁻⁵ to 10⁻² M; phosphate or acetate buffer = 0.016 M; the theoretical curve was calculated from Eq. 10. The constants were determined by trial and error to be k_r' = 27 [M⁻³ sec.⁻¹], k_r'' = 3 × 10⁻³ [M⁻² sec.⁻¹], Ka = 3 × 10⁻³ [M] at 25.0°.

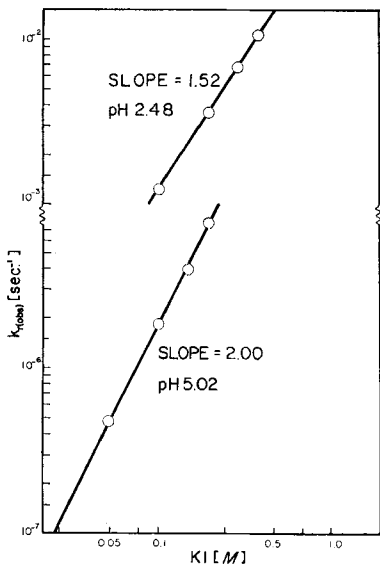


Fig. 4.—Log-log plot of observed first-order rate constant vs. potassium iodide for the reduction of dehydromethionine. At pH 5.02: succinate buffer = 0.1 M; C₅H₉NO₂S = 0.01 M; ionic strength = 0.35 (adjusted with KCl). At pH 2.48: phosphate buffer = 0.01 M; C₅H₉NO₂S = 5 × 10⁻⁵ M; ionic strength = 0.41 (adjusted with KCl); temperature = 25.0°.

neutral species requires two, the following rate law can be written:

$$\frac{d[I_2]_T}{dt} = k_{r(\text{obs.})} [De]_T = k_r' [De] [H^+] [I^-]^2 + k_r'' [DeH^+] [H^+] [I^-] \quad (\text{Eq. 8})$$

where [De]_T, [De], and [DeH⁺] represent the concentration of the total, unprotonated, and protonated dehydromethionine, respectively. The constants k_r' and k_r'' are fourth and third-order rate constants, respectively, for these different mechanisms. Since the dissociation constant of the protonated dehydromethionine is

$$K_a = \frac{[De] [H^+]}{[DeH^+]} \quad (\text{Eq. 9})$$

from Eq. 8 the following rate law can be derived:

$$k_{r(\text{obs.})} = \frac{k_r' K_a [H^+] [I^-]^2}{[H^+] + K_a} + \frac{k_r'' [H^+]^2 [I^-]}{[H^+] + K_a} \quad (\text{Eq. 10})$$

The constant k_r' can be easily determined from the pH profile in Fig. 3, the apparent pseudo first-order rate constant being, above pH 4.5,

$$k_{r(\text{obs.})} \cong k_r' [H^+] [I^-]^2 \quad (\text{Eq. 11})$$

A mean value of k_r', the rate constant for the reverse reaction in essentially neutral system, was calculated on this basis to be 27 [M⁻³ sec.⁻¹] at 25°.

From the shape of the pH profile and from Eq. 10, we can estimate the dissociation constant of the protonated dehydromethionine and the rate constant for the reverse reaction in acid solution to be K_a = 3 × 10⁻³ [M¹] and k_r'' = 7 [M⁻² sec.⁻¹].

Although the dissociation constant was not measured directly, the obtained pK_a value of 2.52 appears reasonable on another basis. From the structure point of view, a pK_a value not too different from that of methionine (pK_a = 2.22) appears to be likely because the positive sulfur in dehydromethionine may be expected to increase the dissociation of the carboxyl group as the protonated amino group in methionine does. Furthermore, the pH profile of the hydrolysis of dehydromethionine to methionine sulfoxide (Fig. 10), as will be discussed later, corresponds to the same value based on the assumption that the protonated dehydromethionine hydrolyzes with a different rate than the neutral species.

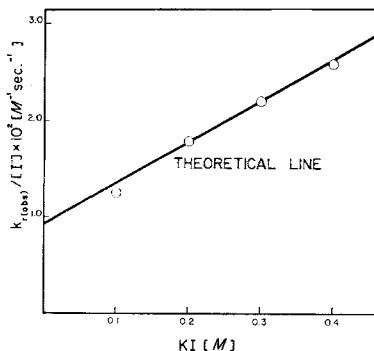


Fig. 5.—According to Eq. 12, observed first-order rate constants obtained at pH 2.48 (see Fig. 4) for the reduction of dehydromethionine divided by the iodide concentration are shown plotted against the iodide concentration. The theoretical curve was calculated from Eq. 12 using the constants showing best fit for pH profile in Fig. 3.

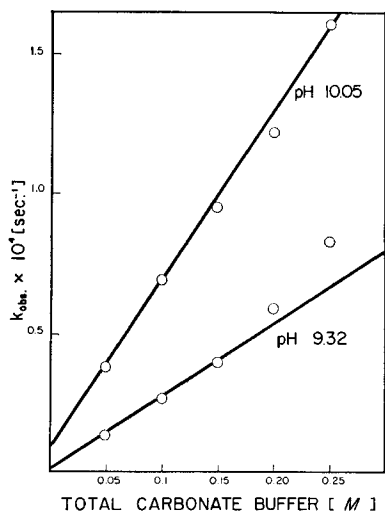


Fig. 8.—Observed first-order rate constants vs. total carbonate buffer concentration for the hydrolysis of dehydromethionine (measured titrimetrically). Buffer ratio = 0.3 mole HCO_3^- /0.7 mole CO_3^{2-} at pH 10.05; 0.7 mole HCO_3^- /0.3 mole CO_3^{2-} at pH 9.32. $\text{C}_5\text{H}_9\text{NO}_2\text{S} = 0.01 M$; ionic strength = 0.875 (adjusted with KCl); temperature = 2.50° .

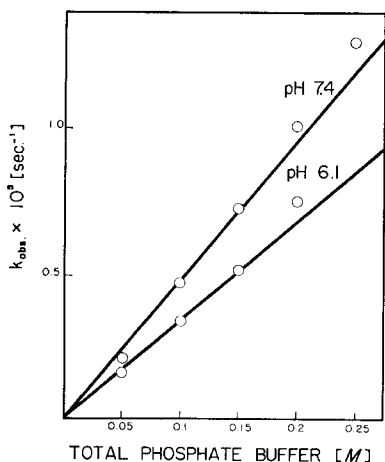


Fig. 9.—Observed first-order rate constants vs. total phosphate buffer concentration for the hydrolysis of dehydromethionine (measured titrimetrically). Buffer ratio: 0.15 mole H_2PO_4^- /0.85 mole HPO_4^{2-} at pH 7.4; 0.7 mole H_2PO_4^- /0.3 mole HPO_4^{2-} at pH 6.1. $\text{C}_5\text{H}_9\text{NO}_2\text{S} = 0.01 M$; ionic strength = 0.875 (adjusted with KCl); temperature = 25.0° .

straight lines from which observed pseudo first-order rate constants could be calculated. In strongly acidic solutions, the rate was also measured spectrophotometrically by following the decrease in absorbance at 243 $m\mu$.

Figures 7, 8, and 9 show the influence of the different buffer species on the rate of hydrolysis of dehydromethionine. In these graphs the observed pseudo first-order rate constants obtained in presence of acetate, carbonate, and phosphate buffer are

shown plotted against the total buffer concentration. For most plots the experimental points fall essentially on a straight line, the intercepts representing pseudo first-order rate constants at the particular pH in absence of any buffer and the slopes the specific second-order rate constants for the buffer species involved.

$$k_{\text{obs.}} = k_{\text{H}_2\text{O}} + k_{\text{T}} [\text{buffer}]_{\text{T}} \quad (\text{Eq. 13})$$

Since each buffer mixture consisted of an acid and a basic species, the following equation for the slope can be written

$$k_{\text{T}} = k_{\text{base}} \times F_{\text{base}} + k_{\text{acid}} \times F_{\text{acid}} \quad (\text{Eq. 14})$$

where F represents the fraction of the basic or acidic species, *i.e.*, $F_{\text{base}} + F_{\text{acid}} = 1$. Since for each buffer system two series of experiments with different buffer ratios were carried out from the slopes of these two plots and the particular fractions, the catalytic second-order rate constant for the acidic species and the basic species was calculated. These rate constants are summarized in Table I.

The rate of hydrolysis in absence of any buffer has also been studied, mainly at low and at high pH, the hydrogen-ion concentration was kept constant by repeatedly adding sodium hydroxide solution. The results of these studies and the extrapolated values for zero buffer concentration of Figs. 7, 8, and 9 are presented as the pH profile in Fig. 10 showing that dehydromethionine hydrolyzes easily in presence of hydrogen ions and hydroxide ions and is very stable at neutral pH ($t_{1/2} \approx 600$ days).

As can be seen in Fig. 10, the rate of hydrolysis in strong acid solution is about only a third of the rate which could be expected by extrapolation from the observed values at pH 5. This decrease of the observed rate constants may be due to the slower rate of reaction of the protonated species relative to the free form. On the basis of this assumption, the following rate law for the hydrolysis of dehydromethionine in acidic solution and in the absence of any buffer species can be written:

$$\frac{-d[\text{De}]_{\text{T}}}{dt} = k_{\text{obs.}} [\text{De}]_{\text{T}} = k_{\text{H}^+} [\text{De}] [\text{H}^+] + k'_{\text{H}^+} [\text{DeH}^+] [\text{H}^+] \quad (\text{Eq. 15})$$

Considering Eq. 9, the observed first-order rate constant can be expressed as

$$k_{\text{obs.}} = \frac{k_{\text{H}^+} K_{\text{a}} [\text{H}^+] + k'_{\text{H}^+} [\text{H}^+]^2}{[\text{H}^+] + K_{\text{a}}} \quad (\text{Eq. 16})$$

From Eq. 16 and the experimental data of Fig. 10, the constants were determined to be $k_{\text{H}^+} = 6 \times 10^{-2} [M^{-1} \text{sec.}^{-1}]$, $k'_{\text{H}^+} = 1.7 \times 10^{-2} [M^{-1} \text{sec.}^{-1}]$, and $K_{\text{a}} = 3 \times 10^{-3} [M]$ which agrees with the value obtained from Fig. 3. The theoretical curve of

TABLE I.—CATALYTIC CONSTANT FOR HYDROLYSIS OF DEHYDROMETHIONINE FOR SEVERAL BUFFER SPECIES FROM FIGS. 7, 8, AND 9 AT 25°

Catalytic Active Species	$k [M^{-1} \text{sec.}^{-1}] \times 10^4$
CH_3COOH	1.31
CH_3COO^-	0.0013
H_2PO_4^-	0.25
HPO_4^{2-}	0.5
HCO_3^-	0.09
CO_3^{2-}	8.6

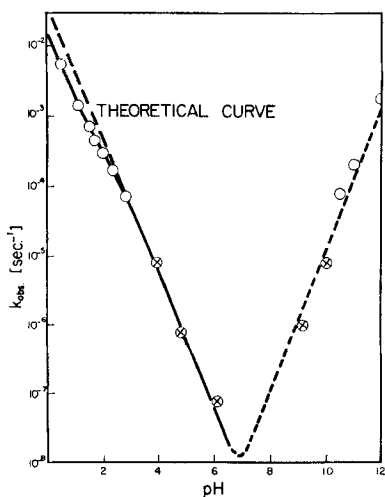


Fig. 10.—Observed first-order rate constants *vs.* pH for the hydrolysis of dehydromethionine (measured titrimetrically and spectrophotometrically). Key: O, determined directly; ⊗, determined by extrapolation from Figs. 7, 8, and 9. $C_6H_9NO_2S = 0.01 M$; ionic strength = 0.875 (adjusted with KCl); temperature = 25.0°. The theoretical curve was calculated from Eq. 16, derived from the assumption of different rates for unprotonated and protonated dehydromethionine, values of k_H^+ , k'_H^+ , and K_a of Eq. 16 were chosen to fit best the experimental points.

Fig. 10 is calculated from Eq. 16 and is in good agreement with the experimental results.

The mechanism of the hydrolysis of dehydromethionine in alkali solution seems to be best rationalized by assuming a nucleophilic attack of hydroxide ion or anionic buffer species on the positive sulfur, followed by ring opening and protonation of the nitrogen. In acidic solution protonation of the imino group probably occurs first, followed by a water attack on sulfur. The relatively high catalytic activity of acetic acid compared to the acetate species might be caused through a nucleophilic attack of the carbonyl oxygen on the sulfur. Protonation of the imino group occurs then through general acid catalysis of the attached carboxyl group.

The influence of iodide ions on the rate of hydrolysis of dehydromethionine as mentioned by Lavine (1) was not observed. It seems to be likely that Lavine was not aware of the catalytic activity of phosphate buffer.

Application of Kinetics Leading to Improvement of Assay Method.—In the original method proposed by Lavine, dehydromethionine stands for 20 min. in contact with 0.2 *M* phosphate buffer at pH 7. It is evident from the data given above on the hydrolysis of dehydromethionine that a significant portion will be observed to form a sulfoxide species which would not undergo reversible reduction back to the amino acid. The extent of loss on the basis of our experiment would be of the order of 1%. Another small error of about 0.2% in the presence of 1 *M* potassium iodide solution and at pH 1 can be calculated from Eqs. 10 and 16 because

the reduction and the hydrolysis of dehydromethionine are competitive reactions.

From the relative high value of the specific second-order rate constant $k_f = 6.5 \times 10^6 [M^{-1} \text{ sec}^{-1}]$ of the forward reaction, it could be expected that at about pH 9 and in presence of low potassium iodide concentration methionine can be directly titrated with iodine. This possibility has already been mentioned by Lavine (1); no result, however, has been presented. The following procedure was tested with the same lot of methionine.

About 150 mg. methionine, previously dried at 60° in vacuum for 2 hr., was accurately weighed and dissolved in 50 ml. of water, with 1 Gm. of sodium borate. After addition of 1 ml. of 1% starch solution, it was titrated with 0.1 *N* iodine until a slightly blue color remained for at least 1 min.

The results are given in Table II for 9 separate analyses and compared with 9 independent determinations of the same sample according to the procedure of N.F. XII.

Both methods have practically the same precision judged by the standard deviation. The differences of about 0.5% between the mean values seems to be partially due to the different ways by which the end point is approached. In the direct iodometric titration a slight excess of iodine is added to produce the starch-iodine complex, while for the back titration of iodine with sodium thiosulfate, a slight excess of the titrant is unavoidable. Therefore, the difference of about 0.5% includes the sum of the titration errors of the two methods. Both methods were also carried out in the presence of glycine (equivalent to 10% of the applied amount of methionine). No higher values were observed.

The value obtained by direct titration can also be checked by continuing with an indirect determination of methionine in the same solution.

The slight excess of iodine is carefully removed by sodium thiosulfate solution, 12 Gm. of potassium iodide and 10 ml. of 25% hydrochloric acid are added, and the liberated iodine is titrated with 0.1 *N* sodium thiosulfate solution. Since in a solution of 1 *M* potassium iodide at low pH the iodide species can easily be oxidized by air, a blank is performed by preparing a solution of 1 Gm. sodium borate, 11 Gm. potassium iodide, and 10 ml. of 25% hydrochloric acid in 70 ml. of water and titrating the liberated iodine after 1 to 2 min. of stirring.

The mean value of the indirect determination and its standard deviation is also given in Table II and shows very good agreement with the direct titration. Since this indirect procedure, however, shows no advantages, the direct iodometric method appears to be sufficient and also superior to the N.F. XII assay, because it can be carried out in much shorter time.

TABLE II.—RESULTS OF A NEW DIRECT AND INDIRECT IODOMETRIC METHOD OF ASSAY OF METHIONINE COMPARED WITH THE DETERMINATION IN "THE NATIONAL FORMULARY," 12TH ED.

Procedure	No. of Determinations	Mean Value, %	S. D., %
Direct	9	100.17	±0.22
Indirect	9	100.00	±0.25
N.F. XII	9	99.68	±0.24

EXPERIMENTAL

Reagents.—Methionine (Eastman-Kodak, white label) was recrystallized from 40% ethanol. All other reagents were analytical grade. Water was purified by distilling tap distilled water from a potassium permanganate solution acidified with sulfuric acid.

The rate of oxidation of methionine with iodine and the rate of reduction of dehydromethionine by iodide forming iodine have been determined spectrophotometrically with a Cary model 11 or 15 by measuring the change in absorbance at 353 μ , the absorption peak of I_3^- .

For the forward reaction, in general 5 ml. of a methionine solution was added to 45 ml. of a solution of buffer, potassium iodide, and iodine to yield 50 ml. of a solution with concentration as given for each figure. Very fast reactions were measured by injecting 1 ml. of methionine solution through the cover of the cell compartment into 2 ml. of solution containing buffer, iodide, and iodine while the instrument was operating.

For studying the reverse reaction, a stock solution of 0.025 *M* dehydromethionine was prepared by adding an equivalent amount of iodine and sodium hydroxide solution to a methionine solution, the pH was kept between pH 5 and 7. Iodide was removed with silver nitrate, the filtrate was brought to the calculated volume. Pseudo first-order rate constants were obtained as described above in the section for the reverse reaction.

The rate of hydrolysis of dehydromethionine was determined in 0.01 *M* solution of the cyclic species in the presence and in the absence of the buffer species.

In both cases 10-ml. samples were withdrawn from the thermostated stock solution, 3 Gm. potassium iodide and 10 ml. 0.5 *N* hydrochloric acid were added, and the liberated iodine was titrated with 0.01 *N* sodium thiosulfate solution. From a plot of log (ml. 0.01 *N* $Na_2S_2O_3$) versus time, pseudo first-order rate constants were obtained.

Since dehydromethionine hydrolyzes under acid formation, the rate of hydrolysis in the absence of any buffer was carried out under simultaneous addition of sodium hydroxide solution. For this purpose the reaction solution was kept in a water jacket container closed with a ground-glass plate. Through the holes in the glass plate, electrodes and the tip of a microburet filled with 1 *N* sodium hydroxide solution were immersed into the solution stirred by a magnetic stirrer. Since for most cases a half-life of more than 10 min. was expected, the pH could be easily controlled manually.

In strong acidic solution the rate of hydrolysis was also determined spectrophotometrically by following a decrease in absorbance at 243 μ .

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Coumarins IV

Coumarins of *Pteryxia terebinthina*. Structures of Two New Coumarins, Isopteryxin and Calipteryxin

By B. EICHSTEDT NIELSEN* and T. O. SOINE

The ether extract of the root of *Pteryxia terebinthina* var. *californica* (Coul. and Rose) Mathias, in addition to osthol, anomalin (I), and pteryxin (III), afforded two new coumarins: 3'(*S*),4'(*S*)-3'-angeloyloxy-4'-acetoxy-3',4'-dihydroseselin (IV) and 3'(*S*),4'(*S*)-3'-angeloyloxy-4'-seneciolyloxy-3',4'-dihydroseselin (V). These coumarins are provisionally named isopteryxin and calipteryxin, respectively.

IN KEEPING with an interest in naturally occurring coumarins (1) and prompted by the recent isolation of anomalin (I) from *Angelica*

anomala Lall by Hata *et al.*, and its structure determination as the 3',4'-diangelate ester of 3',4'-dihydroxy-3',4'-dihydroseselin (II), the present paper reports the isolation of I and several related coumarins from *Pteryxia terebinthina* var. *californica* (Coul. and Rose) Mathias.¹ In particu-

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¹ The authors are indebted to Dr. T. G. Call, California State Polytechnic College, San Luis Obispo, Calif., for collection and identification of the plant material. Preliminary experiments by Dr. Call had shown the presence of osthol and a new coumarin which has since been identified as isopteryxin. Suitable specimens of the plant material have been placed in the herbarium of the Botany Department, University of Minnesota.